

Blood Collection Tip Sheet

Order of draw:

1. Blood Cultures
2. Blue
3. Gold/Red
4. Green
5. Purple
6. Gray

Preventing Hemolysis

- Mix tubes with anticoagulant additives gently 5-10 times.
- Avoid drawing blood from a hematoma.
- Avoid drawing the plunger back too forcefully.
- Make sure venipuncture site is dry.
- Avoid probing.
- Avoid prolonged tourniquet application or fist clenching, tourniquets should not be on longer than one minute.

Preventing Clotting

- By gently inverting the tube 5-10 times, the blood will mix with anticoagulant additive and clotting should not occur.
- Delay transferring from a syringe into tubes
- Avoid using expired tubes.
- Avoid overfilling tubes (forcing blood with a syringe).

Patient Identification

- Verify patient's full name and date of birth; Compare patient's full name and date of birth with barcode label.
- If the patient is unresponsive, verify identification via armband.
- Samples must be labeled, and collected at the patient's bedside utilizing beaker collection process. They should NEVER leave the room unlabeled.
- Only send one patient's samples per specimen bag.

Blood Bank Specimens

- Patient identity must be verified by two individuals, the transfusionist and a qualified witness.
- **DO NOT TRANSFUSE** if any discrepancies are found during this identification process and immediately return the product to the Blood Bank for resolution.
- An electronic signature is required. Sample will be rejected if this step is not completed properly.
- Blood Bank tubes can NEVER be relabeled.

Collection of Blue Top Tubes

- When using a winged blood collection set for venipuncture and a blue (coagulation) tube is the first specimen to be drawn, a discard tube should be drawn first.
- The discard tubes is used to fill the blood collection's tubing "dead space."
- The discard tube does not need to be completely filled.
- The discard tube should be a blue top tube.

Blood Culture Collection

- Never cover the factory label, the analyzer reads this barcode to obtain the bottle type (aerobic or anaerobic).
- You MUST specify the collection site.
- Use the Steripath when collecting samples.

- Apply a Beaker label to each blood culture bottle, taking care NOT TO COVER the areas shown below in RED, including:



Phlebotomy Rounds

- Orders with a priority of **ROUTINE** will be drawn +/- 1 hours of the collect time. (For example, an order for 1400 will be collected between 1300 and 1500).
- Orders with a priority of **TIMED** and **ASAP** will be drawn +/- 1/2 hour of the collect time. (For example, an order for 1500 will be collected between 1430 and 1530).

Miscellaneous

- Always mix tubes directly after collection, this will help mix anticoagulant with blood to prevent clotting.
- **Inappropriate Sites for Venipuncture:** Arm on side of mastectomy, Edematous areas, Hematomas, Scarred areas, Arms with fistulas or vascular grafts and Sites above an IV cannula.
- Blood should not be collected during a transfusion

Blood Collection Tip Sheet

Quantiferon Gold Plus (QTB) Collection and Processing Instructions

Note: QTB specimens that have not been incubated and centrifuged will be rejected. See steps 4-7.

1. Collect 1 mL of blood into **each of 4 tubes**.
 - **Tubes fill slowly** **Order of Draw: Nil, TB1, TB2, Mitogen (Gray, Green, Yellow, Purple)**
 - Use of a syringe may insure correct blood volume
 - When tube is upright, blood volume must be within the small black mark on the label.
 - If butterfly needle is used, first collect other required tubes or use a “purge” tube to remove the air and then proceed with collecting the QTB tubes.
2. Immediately **shake** the tubes firmly for **5 seconds** (10x)
 - Entire inner surface of tube must be coated with blood.
 - Over-energetic shaking may cause gel disruption and could lead to aberrant results.
3. Label tubes such that “Quantiferon” band and entire specimen volume are visible.
4. Four labels will be printed and labels must be placed on correct tube based on the cap color matching the color stated on each barcode label.
5. Maintain tubes at room temperature until incubation.
 - **It is recommended that incubation at the facility start as soon as possible.**
 - **Incubation must be started within 16 hours of collection.**
6. Invert tubes for 5 seconds immediately before incubation.
7. **Incubate all 4 tubes upright at 37°C for 16 – 24 hours**
 - **Do Not** use waterbath for incubation.
 - Improper incubation may cause erroneous results
8. **Centrifuge tubes for 15 minutes at 3,000 RCF (g).**
9. Place all 4 tubes together in the QNTF rack in the refrigerator for special chemistry.